
CHEMICAL COMPOSITION AND ANTIFUNGAL ACTIVITY OF FOUR ESSENTIAL OILS AGAINST PHYTOPATHOGENS RESPONSIBLE FOR ROOT ROT OF WHEAT IN MOROCCO

El Mostafa Zahraoui^{a,b}, Bouchra Ababou^a, Brahim El Yousfi^b, Khadija Boukachabine^{a,*}

^aLaboratory of Environmental Sciences and Development, University Hassan 1st, Faculty of Sciences and Technology, Settat, Morocco

^b Laboratory of Phytopathology, Regional Center of Agronomic Research, Settat, Morocco

* Corresponding Author:

Khadija Boukachabine

Phone: (00212) 5 23 40 07 36, Fax : (00212) 5 23 40 09 69

ABSTRACT

The essential oil (EO) isolated by hydro-distillation from the leaves, stems and flowers of *Thymus satureoides*, *Origanum compactum*, *Mentha pulegium* and *Artemisia herba alba* from Moroccan plants were analyzed by GC–MS. The major components were respectively borneol (26,45%) and thymol (12,79%) of EO from *Thymus satureoides*, carvacrol (43,97%) and p-Cymene (17,87%) of EO from *Origanum compactum*, pulegone (78,07%) of EO from *Mentha pulegium* and thujone (59,07%) and α -campholene aldehyde (12,71%) of EO from *Artemisia herba alba* L. Antifungal activity of these four oils were tested by poisoned food technique against two plant pathogenic fungi viz *Bipolaris sorokiniana* (Bs) and four *Fusarium culmorum*'s isolates (Fc1, Fc2, Fc3 and Fc4). Minimum inhibitory concentration (MIC), minimum fungi static concentration (MFC) and median inhibitory concentration (IC50) for Isolates tested showed a highly significant variability in their reaction against the action of essential oils tested. Isolate Fc2 was the most resistant to the action of the four essential oils compared to other strains, with IC50 ranging between 0.147 and 4.496mg/mL, MIC from 0.29 to 9.11mg/mL and MFC between 0.57 and 9.11mg/mL. These results suggested that these essential oils could provide natural alternatives against Fc and Bs.

Keywords: *Fusarium culmorum*; *Bipolaris sorokiniana*; Wheat; Essential oils; antifungal activity.

1. Introduction

Crop losses due to fungal diseases can amount to 12% in developing countries (Bajpai *et al.*, 2008). Fungal diseases caused by soil fungi induce a significant loss in yield and quality of crops (Kumar *et al.*, 2014). In Morocco, Fc and Bs cause root rot, common root, spot blotch and scald in cereal crops (Murray and Brennan, 2009; Chekali *et al.*, 2011). Frequent using of wide variety of synthetic chemical compounds of high concentrations as antimicrobial agents against phytopathogenic fungi in agriculture (Bajpai *et al.*, 2008) and prevalence of resistant pathogenic strains (Chebli *et al.*, 2003).

The EO preparations that possess antimicrobial activities (Riccioni and Orzali, 2011) have been the subject of many investigations (Matasyoh *et al.*, 2007). Again, EOs of some plants have recently been proven to be successful bio-control agent (Chutia *et al.*, 2006 ; Sokovic and Griensven, 2006). It was observed these EOs from *Origanum compactum* against foodborne bacteria (Sbayou *et al.*, 2014a), *Mentha pulegium* and *Artemisia herba alba* strongly inhibited bacterial growth (Sbayou *et al.*, 2014b). But there is no report of the antifungal activity of these Eos in phytopathogenic fungi. Therefore, the present study was made to determine antifungal activity of these EOs against some phytopathogenic fungal species viz Fc and Bs, with emphasis on the possible future application of these EOs as alternative antifungal agents.

2. Material and Methods

2.1. Fungal strains

Five fungal isolates responsible for root rot of durum wheat crops were isolated from plant pathology (Agronomic Research Moroccan Institutes). Four isolates of *Fc* (Fc1, Fc2, Fc3 and Fc4) and one isolate of *Bs* were tested.

2.2. Essential oils and chemical composition

EOs were extracted by steam distillation from the leaves, stems and flowers from Moroccan plant *Thymus satureoides*, *Origanum compactum*, *Mentha pulegium* and *Artemisia herba alba* (SANTIS-sarl Company in Morocco). Analysis of the chemical composition was performed by gas chromatography (GC Ultra Trace) coupled to a mass spectrometer (MS Polaris Q ion trap). GC is equipped with a VB-5 column (5% phenyl methylpolysiloxane), 30m*0.25mm*0.25µm for detecting an electron ionization system with 70eV was used. The carrier gas was helium with a flow rate of 1.4 mL/min. Two molecules of the triazoles family (Tetraconazole and Hexaconazole), widely used in the fungal disease control (Chekali *et al.*, 2011), are used as positives controls.

2.3. Determination of IC₅₀, MFC and MIC

The evaluation of inhibition percentage of EOs tested was performed using the technique reported by Soylu *and Soylu* (2010). IC₅₀ was determined from the equations of dose-response curves of concentrations of essential oils tested (Kumar *et al.*, 2014). Minimum fungicidal concentrations (MFC) were determined by transferring the mycelium disc, which have not grown, on the PDA medium without essential oil (Soylu *and Soylu*, 2010). IC₅₀ and MFC values obtained were the median of three replicates. The results of the average of three replicates were compared by analysis of variance (ANOVA) followed by comparison of means by the Duncan test at a probability of 5% by SPSS 22 software.

3. Results

Chromatographic profiles and mass spectra show that the four EOs have different chemical compositions; the major compounds vary according to each essential oil (Table 1). Carvacrol represents 43.97% of the total composition of the essential oil of *Origanum compactum*, followed by p-cymene (17.87%), which is a precursor of thymol and carvacrol, thymol (11.56%) and γ -terpinene (8.88%). Borneol (26.45%) and thymol (12.79%) were the major compounds of the *Thymus satureoides* essential oil. Pulegone composes 78.07% of essential oil of *Mentha pulegium*. Thujone is the major compound (59.07%) from essential oil chemical composition of *Artemisia herba alba* analyzed in this study, followed by α -campholene aldehyde.

Table 1: Chemical composition of essential oils tested				
Compounds	Percentage of compounds			
	<i>Origanum compactum</i>	<i>Thymus satureoides</i>	<i>Mentha pulegium</i>	<i>Artemisia herba alba</i>
Carvacrol	43.97	1.51	-	-
p-Cymene	17.87	2.18	-	-
Pulegone	-	-	78.07	-
Thymol	11.56	12.79	-	-
Thujone	-	-	-	59.07
γ -Terpinene	8.88	1.61	-	0.32
Dodecamethyl-1,11-dihydrohexasiloxane	8.71	8.47	13.43	5.26
α -Campholene	-	-	-	12.71

aldehyde				
Caryophyllene	1.85	9.17	1.89	-
Carveol (fr.1)	1.8	-	-	-
α -Terpinene	1.15	0.5	-	-
α -Thujone	-	-	-	11.73
α -Pinene	1.05	5.19	0.70	1.02
Borneol	0.23	26.45	-	-
α -Terpinenyl acetate	-	10.99	-	-
Camphene	-	7.16	-	-
Santolina triene	-	1.93	0.8	-
Endobornyl acetate	-	1.87	-	-
Alloocimene	-	1.56	-	-
Artemisia triene	-	-	-	3.75
β -Cadinene	-	1.41	-	-
γ -Cadinene	-	1.35	-	-
Copaene	-	0.37	-	-
Isopulegone	-	-	1.45	-
p-Menthone	-	-	1.1	-
α -Cedrol	-	-	0.51	-
Sabinene	-	-	-	3.05
Germacrene-D	-	-	-	0.44
Patchoulane	-	-	-	0.3
Nopol	-	-	-	0.72
Cis-Sabinene hydrate	-	-	-	0.93
α -Muurolene	-	-	-	0.16
Thujol	-	-	-	0.13
Total	97.07	97.18	97.95	99.59

Significant differences are observed between EOs tested and positives controls (Table 2). IC₅₀ values show that the essential oil of *Origanum compactum* exerts the most important antifungal activity compared to other EOs tested and positives controls, between 0.018mg/mL and 0.147mg/mL. The significant antifungal activity of this EO is due to the presence of the phenolic compounds carvacrol and thymol. EOs of *Mentha pulegium* and *Thymus satureoides* have an average antifungal activity, compared to other essential oils tested. Antifungal activity

from EO of *Mentha pulegium* is represented by IC₅₀ ranging of 0.034mg/mL to 0.578mg/mL and between 0.154mg/mL and 0.582mg/mL for EO of *Thymus satureoides*. EO of *Mentha pulegium* was more active than *Thymus satureoides*. EO of *Artemisia herba alba* exerts, compared to other essential oils, has lowest action against isolates tested, with IC₅₀ ranging from 0.725mg/mL to 4.496mg/mL.

Isolates tested showed a highly significant variability in their reaction against the action of EOs tested. Fc2 isolate resist to the action of the four essential oils tested with higher IC₅₀, MIC and MFC compared to other strains, IC₅₀ between 0.147 and 4.496mg/mL, MIC ranging from 0.29 to 9.11mg/mL and MFC between 0.57 and 9.11mg/mL (Table 2). Fc4 isolate is less resistant than Fc2 to the antifungal action of the three essential oils, *Origanum compactum*, *Mentha pulegium* and *Thymus satureoides* while the most sensitive isolates are Fc1, Fc3 and Bs strain. Whereas the strain of Bs is less resistant than Fc2 isolate to the essential oil of *Artemisia herba alba*. Fc3 and Fc4 isolates are more sensitive to the action of this essential oil (Table 2).

Table 2 : IC ₅₀ , MIC and MFC (mg/mL) of EOs tested and positives controls															
	Fungal isolates														
	Fc1			Fc3			Fc2			Fc4			Bs		
	IC ₅₀	MIC	MFC	IC ₅₀	MIC	MFC	IC ₅₀	MIC	MFC	IC ₅₀	MIC	MFC	IC ₅₀	MIC	MFC
Essential oils															
<i>Origanum compactum</i>	0.018 _a	0,07 ^a	0.29 ^a	0.04 _{4^a}	0,14 ^a	0.14 ^a	0.14 _{7^a}	0,29 ^a	0.57 ^a	0.07 _{4^a}	0,14 ^a	0.29 ^a	0.04 _{6^a}	0,14 ^a	0.29 ^a
<i>Thymus satureoides</i>	0.172 _c	0,29 ^b	1.14 ^b	0.32 _{5^c}	0,57 ^b	1.14 ^b	0.58 _{2^b}	1,14 ^b	2.28 ^c	0.38 _{2^b}	0,57 ^b	1.14 ^b	0.15 _{4^b}	0,29 ^a	1.14 ^b
<i>Mentha pulegium</i>	0.071 _b	0,57 ^b	1.14 ^b	0.16 _{9^b}	1,14 ^c	1.14 ^b	0.57 _{8^b}	2,28 ^c	4.56 ^d	0.32 _{7^b}	1,14 ^c	1.14 ^b	0.03 _{4^a}	1,14 ^c	1.14 ^b
<i>Artemisia herba alba</i>	1.9 ^f	2,28 ^d	4.56 ^c	0.72 _{5^d}	1,14 ^c	1.14 ^b	4.49 _{6^d}	9,11 ^d	>9.1 ₁	0.77 _{7^c}	1,14 ^c	2.28 ^c	2.28 ^d	4,56 ^d	>9.1 ₁
Positive controls															
Triazoles															
- <i>Hexaconazole</i>	0.25 ^d	0.5 ^b	0.5 ^a	0.25 ^c	0.5 ^b	0.5 ^a	0.25 ^a	0.5 ^a	0.5 ^a	0.25 ^b	0.5 ^b	0.5 ^a	0.25 ^b	0.5 ^b	0.5 ^a
- <i>Tetraconazole</i>	0.625 _e	1.25 ^c	1.25 ^b	0.62 _{5^d}	1.25 ^c	1.25 ^b	0.62 _{5^b}	1.25 ^b	1.25 ^b	0.62 _{5^c}	1.25 ^c	1.25 ^b	0.62 _{5^c}	1.25 ^c	1.25 ^b
NB. The values in the same column followed by the same letter are not significantly different according to Duncan test at 5% probability.															

Statistical analysis of IC₅₀ shows no significant difference between the two isolates Fc2 and Fc4 towards essential oils of *Thymus saturooides* and *Mentha pulegium* while this difference is significant between the two other essential oils. IC₅₀ values of *Origanum compactum* are no significant difference with *Mentha pulegium* EO for strain Bs, which exert the most important antifungal action on this strain compared to the other essential oils and positive controls used.

4. Discussion

Chemical composition of the EOs tested show that carvacrol, p-cymene, thymol and γ -terpinene were the major compounds of *Origanum compactum* EO, with some difference in the percentages of compounds (Chebli *et al.*, 2003; Oussalah *et al.*, 2006; Babili *et al.*, 2011) from several Moroccan regions. Borneol and thymol were the major compounds of the *Thymus saturooides* EO, and have been reported as major compounds of Moroccan EOs (Oussalah *et al.*, 2007;

Kloucek *et al.*, 2012). Other chemotypes of Moroccan *Thymus saturooides* have carvacrol as major compound (El Bouzidi *et al.*, 2013). Pulegone is a major compound of EO of *Mentha pulegium* as like plant from north of Morocco (Chraïbi *et al.*, 2016; Ait-Ouazzou *et al.*, 2012) and contrary from Iran (Aghel *et al.*, 2004; Mahboudi *et al.*, 2008). Thujone and α -campholene aldehyde showed high percentages in the *Artemisia herba alba* EO. EO of *Artemisia herba alba* from Tunisian arid zone has also a chemical composition rich with Thujone (Mighri *et al.*, 2010). Geographical origin of the raw material affects the chemical composition of EO (Oussalah *et al.*, 2006; Al-Reza *et al.*, 2010; Paolini *et al.*, 2010). This composition can be affected by weather conditions, soil, species used and extraction technique (Szumny *et al.*, 2010; Russo *et al.*, 2013).

Antifungal activity showed a high significant difference between four EOs tested. The most important antifungal activity was obtained by *Origanum compactum* EO rich in phenolic compounds like carvacrol and thymol. These compounds were more active on several *Fusarium* strains (Dambolena *et al.*, 2012; Marei *et al.*, 2012). The synergy between the carvacrol and thymol leads to an improvement of EO action (Campos-Requena *et al.*, 2015). *Thymus saturooides* and *Mentha pulegium* EOs exerts an average antifungal activity; that's corroborate with Zabka *et al.* (2014) and Hajlaoui *et al.* (2009). EO of *Artemisia herba alba* showed the lowest action, rich with thujone and poor in camphor and 1,8-cineol. Mighri *et al.* (2010) reported that camphor and 1,8-cineol are the most likely compounds to inhibit microbial growth.

Toxicity of the phenols containing in these EOs is primarily based on the inactivation of fungal enzymes containing the SH group in their active sites or by disturbing structure of fungal cell membrane (Hu *et al.*, 2017). On other hand, strains tested have shown a significant

variability against action of tested essential oils, the most resistant isolate was Fc2 and Fc4. This resistance would be due to a selection of resistance strains on farmers' fields in which these strains were isolated. This selection would be induced by unreasonable use of chemical fungicides (Singh and Chhatpar, 2011), which is not the case on the two experimental stations where pesticides use respects the recommended doses, according to standards.

5. Conclusion

Four EOs tested *Thymus satureoides*, *Origanum compactum*, *Mentha pulegium* and *Artemisia herba alba*, showed variable antifungal activity towards three isolates of *Fc* and *Bs* strain. *In vitro* results suggest that these EOs can constitute a source of new fungicidal molecules, to be tested in the field to validate their effectiveness in the fight against wheat root rot.

Conflict of Interest

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

We thank SANTIS-sarl Company in Morocco for the essential oil. The authors are most grateful to the University Hassan 1st and the Center of Agronomic Research, Settat in Morocco, which have supported this work.

References

- Aghel, N., Yamini, Y., Hadjiakhoondi, A., & Pourmortazavi SM. (2004). Supercritical carbon dioxide extraction of *Mentha pulegium L.* essential oil. *Talant*, 62, 407–411.
- Ait-Ouazzou, A., Lorán, S., Arakrak, A., Laglaoui, A., Rota, C., Herrera, A., Pagán, R., & Conchello, P. (2012). Evaluation of the chemical composition and antimicrobial activity of *Mentha pulegium*, *Juniperus phoenicea* and *Cyperus longus* essential oils from Morocco. *Food Research International*, 45, 313-319.
- Al-Reza, S.M., Rahman, A., Ahmed, Y., & Kang, S.C. (2010). Inhibition of plant pathogens *in vitro*, *in vivo* with essential oil, and organic extracts of *Cestrum nocturnum L.* *Pesticide Biochemistry and Physiology*, 96(2), 86–92.
- Babili, F., El Bouajila, J., Souchard, J.P., Bertrand, C., Bellvert, F., Fouraste, I., Moulis, C., & Valentin, A. (2011). Oregano: Chemical analysis and evaluation of its antimalarial, antioxidant, and cytotoxic activities. *Journal of Food Science*, 76(3), 512-518.

- Bajpai, V.K., Shukla, S., & Kang, S.C. (2008). Chemical composition and antifungal activity of essential oil and various extract of *Silene armeria* L. *Bioresource Technology*, 99, 8903-8908.
- Campos-Requena, V.H., Rivas, B.L., Pérez, M.A., Figueroa, C.R., & Sanfuentes, E.A. (2015). The synergistic antimicrobial effect of carvacrol and thymol in clay/polymer nanocomposite films over strawberry gray mold. *LWT - Food Science and Technology*, 64 (1), 390-396.
- Chand, R., Singh, H.V., Joshi, A.K., & Duveiller, E. (2002). Physiological and morphological aspect of *Bipolaris sorokiniana* conidia surviving on wheat straw. *The plant pathology journal*, 18(6), 328-332.
- Chebli, B., Mohamed, A., Idrissi Hassani, L.M., & Mohamed, H. (2003). Chemical composition and antifungal activity of essential oils of seven Moroccan Labiatae against *Botrytis cinerea* Pers. *Fr. Journal of Ethnopharmacology*, 89, 165-169.
- Chekali, S., Gargouri, S., Paulitz, T., Nicol, J., Rezgui, M., & Nasraoui, B. (2011). Effects of *Fusarium culmorum* and water stress on durum wheat in Tunisia. *Crop Protection*, 30, 718-725.
- Chraïbi, M., Farah, A., Lebrazi, S., Elamin, O., Houssaini, M.I., & Fikri-Benbrahim, K. (2016). Antimycobacterial natural products from Moroccan medicinal plants: Chemical composition, bacteriostatic and bactericidal profile of *Thymus satureioides* and *Mentha pulegium* essential oils. *Asian Pac J Trop Biomed*, 6(10), 836-840.
- Chutia, M., Mahanta, J.J., Saikia, R.C., Baruah, A.K.S., & Sarma, T.C. (2006). Influence of leaf blight disease on yield of oil and its constituents of *Java citronella* and in-vitro control of the pathogen using essential oils. *World Journal of Agriculture Science*, 2(3), 319–321.
- Dambolena, J.S., López, A.G., Meriles, J.M., Rubinstein, H.R., & Zygadlo, J.A. (2012). Inhibitory effect of 10 natural phenolic compounds on *Fusarium verticillioides*. A structure-property-activity relationship study. *Food Control*, 28(1), 163–170.
- El Bouzidi, L., Jamali, C.A., Bekkouche, K., Hassani, L., Wohlmuth, H., Leach, D., & Abbad, A. (2013). Chemical composition, antioxidant and antimicrobial activities of essential oils

obtained from wild and cultivated Moroccan Thymus species. *Industrial Crops and Products*, 43, 450–456.

Hajlaoui, H., Trabelsi, N., Noumi, E., Snoussi, M., Fallah, H., Ksouri, R., & Bakhrouf, A. (2009). Biological activities of the essential oils and methanol extract of two cultivated mint species (*Mentha longifolia* and *Mentha pulegium*) used in the Tunisian folkloric medicine World. *Journal of Microbiology and Biotechnology*, 25, 2227–2238.

Hu, Y., Zhang, J., Kong, W., Zhao, G., & Yang, M. (2017). Mechanisms of antifungal and anti-aflatoxigenic properties of essential oil derived from turmeric (*Curcuma longa L.*) on *Aspergillus flavus*. *Food Chemistry*, 220, 1–8.

Kloucek, P., Smid, J., Frankova, A., Kokoska, L., Valterova, I., & Pavela, R. (2012). Fast screening method for assessment of antimicrobial activity of essential oils in vapor phase. *Food Research International*, 47(2), 161-165.

Kumar, V., Mathela, C.S., Tewari, G., Darshan, S., Tewari, A.K., & Bisht, K.S. (2014). Chemical composition and antifungal activity of essential oils from three Himalayan Erigeron species. *LWT - Food Science and Technology*, 56, 278-283.

Mahboudi, M., Haghi, G., Mahboubi, M., & Haghi, G. (2008). Antimicrobial activity and chemical composition of *Mentha pulegium L.* essential oil. *Journal of Ethnopharmacology*, 119, 325–327.

Marei, G.I.K., Abdel Rasoul, M.A., & Abdelgaleil, S.A.M. (2012). Comparative antifungal activities and biochemical effects of monoterpenes on plant pathogenic fungi. *Pesticide Biochemistry and Physiology*, 103(1), 56–61.

Matasyoh, J. C., Kiplimo, J. J., Karubiu, N. M., & Hailstorks, T. P. (2007). Chemical composition and antimicrobial activity of essential oil of *Tarchonanthus camphorates*. *Food Chemistry*, 101, 1183–1187.

Mighri, H., Hajlaoui, H., Akrouf, A., Najjaa, H., & Neffati, M. (2010). Antimicrobial and antioxidant activities of *Artemisia herba-alba* essential oil cultivated in Tunisian arid zone. *Comptes Rendus Chimie*, 13(3), 380-386.

Murray, G.M., & Brennan, J.P. (2009). Estimating disease losses to the Australian wheat industry. *Australasian Plant Pathology*, 38(6), 558-570.

- Oussalah, M., Caillet, S., Saucier, L., & Lacroix, M. (2006). Antimicrobial effects of selected plant essential oils on the growth of a *Pseudomonas putida* strain isolated from meat. *Meat Science*, 73 : 236-244.
- Oussalah, M., Caillet, S., Saucier, L., & Lacroix, M. (2007). Inhibitory effects of selected plant essential oils on the growth of four pathogenic bacteria: *E. coli* O157:H7, *Salmonella typhimurium*, *Staphylococcus aureus* and *Listeria monocytogenes*. *Food Control*, 18: 414-420.
- Paolini, J., El Ouariachi, E., Bouyanzer, A., Hammouti, B., Desjobert, J.M., Costa, J., & Muselli, A. (2010). Chemical variability of *Artemisia herba-alba* Asso essential oils from East Morocco. *Chemical Papers*, 64 (5), 550–556.
- Qi, P.F., Johnston, A., Balcerzak, M., Rocheleau, H., Harris, L.J., Long, X.Y., Wei, Y.M., Zheng, Y.L., & Ouellet, T. (2012). Effect of salicylic acid on *Fusarium graminearum*, the major causal agent of *Fusarium* head blight in wheat. *Fungal Biology*, 116, 413-426.
- Riccioni, L., & Orzali, L. (2011). Activity of Tea Tree (*Melaleuca alternifolia*, Cheel) and thyme (*Thymus vulgaris*, Linnaeus). Essential Oils against Some Pathogenic Seed Borne Fungi. *Journal of essential oil research*, 23, 43-47.
- Russo, M., Suraci, F., Postorino, S., Serra, D., Roccotelli, A., & Agosteo, G.E. (2013). Essential oil chemical composition and antifungal effects on *Sclerotium cepivorum* of *Thymus capitatus* wild populations from Calabria, southern Italy. *Revista Brasileira de Farmacognosia – Brazilian Journal of Pharmacognosy*, 23(2), 239–248.
- Sbayou, H., Oubrim, N., Bouchrif, B., Ababou, B., Boukachabine, Kh., & Amghar, S. (2014a). Chemical composition and antibacterial activity of essential oil of *Origanum compactum* against foodborne bacteria. *International Journal of Engineering Research & Technology*, 3 (1), 3562-3567.
- Sbayou, H., Ababou, B., Boukachabine, Kh., Manresa, A., Zerouali, K., & Amghar, S. (2014b). Chemical composition and antibacterial activity of *Artemisia herba-alba* and *Mentha pulegium* essential oils. *Journal of Life Sciences*, 8 (1), 35-41
- Shukla, R., Singh, P., Prakash, B., & Dubey, N.K. (2012). Antifungal, aflatoxin inhibition and antioxidant activity of *Callistemon lanceolatus* (Sm.) Sweet essential oil and its major

component 1,8-cineole against fungal isolates from chickpea seeds. *Food Control*, 25(1), 27-33.

Singh, A.K., & Chhatpar, H.S. (2011). Combined use of *Streptomyces sp.* A6 and chemical fungicides against *Fusarium* wilt of *Cajanus cajan* may reduce the dosage of fungicides required in the field. *Crop Protection*, 30(7), 770–775.

Sokovic, M., & Griensven, L.J.L.D. (2006). Antimicrobial activity of essential oils and their components against the three major pathogens of cultivated button mushroom *Agaricus bisporus*. *European Journal of Plant Pathology*, 116, 211–224.

Soylu, E.M., Kurt, Ş., & Soyly, S. (2010). *In vitro* and *in vivo* antifungal activities of the essential oils of various plants against tomato grey mould disease agent *Botrytis cinerea*. *International Journal of Food Microbiology*, 143, 183-189.

Szumny, A., Figiel, A., Gutiérrez-Ortíz, A., & Carbonell-Barrachina, A. (2010). Composition of rosemary essential oil (*Rosmarinus officinalis*) as affected by drying method. *Journal of Food Engineering*, 97(2), 253–260.

Zabka, M., Pavela, R., & Prokinova, E. (2014). Antifungal activity and chemical composition of twenty essential oils against significant indoor and outdoor toxigenic and aeroallergenic fungi. *Chemosphere*, 112, 443–448.